2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

Impact of the Diet with Purple Sweet Potato and Probiotics on Gas Emission in Swine

Josue David Torres Falcones¹, Jhon Erick Barre Mendoza¹, Ernesto Antonio Hurtado¹

 ${}^{\scriptscriptstyle 1} Veterinary\ Medicine\ Degree, Manuel\ F\'elix\ L\'opez\ Higher\ Polytechnic\ Agricultural\ School\ of\ Manab\'i.\ Calceta,\ Ecuador.$

https://orcid.org/ 0009-0000-6595-2617 https://orcid.org/ 0009-0002-9455-5615 https://orcid.org/0000-0003-2574-1289. ernestohurta@gmail.com

ARTICLE INFO

ABSTRACT

Received: 26 Dec 2024

Revised: 14 Feb 2025

Accepted: 22 Feb 2025

Swine production is one of the main sources of greenhouse gas (GHG) emissions. This study evaluated the impact of including purple sweet potato (Ipomoea batatas L.) and probiotics (Lactobacillus plantarum) in pig diets to mitigate GHG emissions, specifically carbon dioxide (CO₂) and ammonia (NH₃). Thirty-six male pigs were used, distributed into nine dietary treatments combining different proportions of commercial feed, purple sweet potato, and Lactobacillus. CO₂ and NH₃ emissions were measured over 49 days using a gas detector. The results showed that treatments including purple sweet potato and probiotics significantly reduced CO₂ and NH₃ emissions. The most effective treatment combined 40% purple sweet potato with 40 ml of Lactobacillus, achieving a significant reduction in CO₂ (1270 ppm) and NH₃ (24.5 ppm) emissions. The Kruskal-Wallis and Wilcoxon statistical tests confirmed significant differences (P \leq 0.001) between treatments for both variables. In conclusion, the addition of purple sweet potato and probiotics to pig diets is a promising strategy for reducing GHG emissions, contributing to environmental sustainability without compromising the productive performance of pigs.

Keywords: Gas emissions; feed additives; gut microbiota; environmental sustainability; pig diet.

INTRODUCTION

Pig production is one of the main sources of greenhouse gas (GHG) emissions. While providing food for the population, a large amount of waste accumulates, which not only serves as a valuable organic fertilizer but also becomes a source of environmental pollution as gases are released into the atmosphere during decomposition (Vorobel et al., 2024).

It is predicted that emissions of major greenhouse gases will increase by 25% to 90% by 2030 compared to 2000 levels if a series of measures are not adopted to improve the situation (Smith et al., 2008; Udova et al., 2014). The increase in the concentration of several harmful gases (emitted by manure) poses health risks for both animals and humans. The main gases produced on pig farms include methane, hydrogen sulfide, carbon dioxide, ammonia, sulfur dioxide, and volatile fatty acids, which are primarily derived from the fermentation of undigested or poorly digested nutrients (Hossain et al., 2024). In pigs, these gases are produced through microbial fermentation in the large intestine. This complex process is influenced by factors such as diet, intestinal microbiota activity, and digestive physiology, similar to other non-ruminant animals (Rowland et al., 2018).

Given this problem, there is a need to explore sustainable alternatives to reduce greenhouse gas emissions without compromising productive efficiency or animal welfare. In this context, nutritional regulation through dietary modification presents itself as a viable option, especially with the inclusion of feed additives such as probiotics. According to Martínez and Langella (2019), these live microorganisms can provide significant health benefits to the host.

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

Moreover, the polyphenols present in purple sweet potato, when incorporated into the diet, could help reduce the environmental and economic impact associated with its storage and processing, becoming an innovative source of antioxidants (Serra et al., 2021). In addition, the anthocyanins found in this tuber play a key role in promoting growth and health by positively influencing intestinal microbiota, gut physiology, and the immune system (Guo and Shahidi, 2023); besides, Hurtado et al. (2024) indicate the combined supply of sweet potato and *Lactobacillus plantarum* (LP) potentiates the benefits of both ingredients, where sweet potato fiber can help the LP adhere to the intestinal wall and thus improve efficacy.

The rationale for this research lies in the need to find practical and effective solutions to mitigate the environmental impact of pig production. The combination of purple sweet potato and probiotics could offer a novel approach to reducing GHG emissions by leveraging the functional properties of both ingredients to improve digestive efficiency and decrease GHG production. This approach would not only contribute to environmental sustainability but could also enhance the health and performance of pigs, generating economic benefits for producers.

The objective of this study is to evaluate the effect of including purple sweet potato and probiotics in the diet of pigs on the reduction of greenhouse gas emissions. This study aims to provide scientific evidence to support the use of sustainable feeding strategies in pig production, thereby contributing to the reduction of environmental impact through the development of mitigation strategies.

MATERIALS AND METHODS

Ethical Considerations

This study was designed and conducted in strict compliance with international animal welfare and research ethics standards. The experimental protocol, which involved feeding pigs with sweet potato and the inclusion of probiotics to evaluate gas emissions, was reviewed and approved by the Bioethics Committee of the Escuela Superior Politécnica Agropecuaria de Manabí Manuel Félix López (ESPAM MFL). This committee ensures that all research complies with national and international ethical standards concerning the use of experimental animals.

Particular care was taken to ensure that the animals received proper handling, minimizing any form of suffering and providing optimal feeding, housing, and care conditions throughout the experiment. The techniques used to measure gas emissions were non-invasive and did not cause additional stress to the animals.

Animals, diets, and experimental design

Thirty-six crossbred male pigs, resulting from the commercial crossing of sows (Landrace x Duroc) and boars (Pietran), at the fattening stage (100 days old, initial weight 42.77 ± 0.77 kg), were housed in metabolic cages for 49 days. The cages had been previously cleaned, flamed, and disinfected days before the animals' incorporation with a 25% iodine solution.

Commercial fattening feed (CF) was provided according to the nutritional management of the pig herd (ESPAM MFL). The feed had the following composition: crude protein 16%, fat 3%, fiber 4%, ash 8%, and moisture 13%. It was partially replaced with purple sweet potato flour (PSPF) at 20% and 40% inclusion levels (Table 1).

Table 1. Nutritional content of purple sweet potato

Content	Unit/100g
Water	64-74 g
Fiber	1.2-3.5 g
Lipids	0.5-2.1 g
Protein	1.2-7.2 g
Fats	0.4-3.0 g
Carbohydrates	20.19-27.3 g
Sugar	4.18-9.7 g
Glucose	2.37-4.68 mg
Sucrose	56.94-59.97 mg

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

Fructose	1.43-4 mg
Starches	11.8 g

Source: Lim et al., 2016; Wang and Zhu, 2016; Cusumano et al., 2013; Martí et al., 2011; Pagalo et al., 2010; Linares et al., 2008 (cited by Vidal et al., 2018).

The concentration of *Lactobacillus plantarum* (LP) incorporated into the different treatments (1x10¹⁰ CFU.mL⁻¹) was obtained from the molecular biology laboratory (ESPAM MFL) after activation of the corresponding strain.

This study employed a completely randomized experimental design to evaluate the impact of different inclusion levels of purple sweet potato flour and *Lactobacillus* in the diet of pigs on gas emissions, specifically ammonia and carbon dioxide. The primary dependent variable was the amount of these gases emitted, which was measured twice daily (morning and afternoon) over 49 days.

Nine dietary treatments were distributed as follows: 100% CF (T1); 100% CF + 20 mL LP (T2); 100% CF + 40 mL LP (T3); 80% CF + 20% PSPF (T4); 80% CF + 20% PSPF + 20 mL LP (T5); 80% CF + 20% PSPF + 40 mL LP (T6); 60% CF + 40% PSPF (T7); 60% CF + 40% PSPF + 20 mL LP (T8); and 60% CF + 40% PSPF + 40 mL LP (T9).

Each treatment was assigned to two experimental units represented by individual pigs, with two replicates. Gas emission measurements were systematically performed both in the morning and in the afternoon for each animal during two periods (with and without the application of treatments) for 27 and 22 days, respectively.

The completely randomized design employed in this study allowed for the control of intrinsic variability among the animals, ensuring that the differences observed in gas emissions could be reliably attributed to the specific treatments applied. Randomization was key to minimizing experimental bias and enhancing the internal validity of the study, providing robust and scientifically sound results.

Experimental management

The experimental nutritional combinations were provided twice daily: in the morning (8:00 a.m.) and in the afternoon (4:00 p.m.), according to the established treatments. These treatments consisted of partially replacing commercial feed (CF) with purple sweet potato flour (PSPF) and including *Lactobacillus plantarum* (LP).

The determination of the residual gas emission potential from the slurry, composed of a mixture of fresh feces (approximately 50 g) and urine, was carried out using a SKY2000-M6 gas detector. The mixture was placed in a 100 mL laboratory flask, and gas emission measurements (ppm) were conducted daily in two sessions (10:00 a.m. and 5:00 p.m.).

Statistical analysis

The statistical analysis of the relative values (1764 data points) corresponding to gas concentrations (CO₂ and NH₃) was conducted using SAS software (V.9.4, 2023). Since the data did not follow a normal distribution, the non-parametric Kruskal-Wallis test was applied to detect significant differences between treatments. Additionally, descriptive analysis was performed using measures of central tendency (mean) and dispersion (standard deviation, coefficient of variation, maximum, and minimum values) for the different experimental periods (with and without treatment application). The Wilcoxon test for independent samples was used to compare two means, with a significance level set at 0.05.

RESULTS

Descriptive statistics of CO2 and NH3 emissions

Table 2 presents the results from the measurements of CO₂ and NH₃ emissions by pigs under different treatments. The table summarizes the mean concentration values, standard deviation (SD), coefficient of variation (CV), and the minimum and maximum recorded values for both variables.

Table 2. Descriptive statistics of CO2 and NH3 emissions in pigs under different treatments.

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

Treatment	Variable	n	Mean	SD	CV	Min	Max
1	CO ₂	196	1400.00	0.00	0.00%	1400.00	1400.00
NH3		196	30.00	0.00	0.00%	30.00	30.00
2	CO ₂	196	1379.55	17.24	1.25%	1353.00	1400.00
	NH3	196	28.45	1.55	5.45%	26.00	30.00
3	CO ₂	196	1370.13	22.27	1.63%	1340.00	1400.00
	NНз	196	28.26	1.91	6.75%	25.00	30.00
4	CO ₂	196	1359.51	38.63	2.84%	1276.00	1400.00
	NНз	196	28.44	1.53	5.38%	22.00	30.00
5	CO ₂	196	1344.76	39.56	2.94%	1292.00	1400.00
	NH3	196	26.83	3.41	12.71%	22.00	30.00
6	CO ₂	196	1359.53	29.95	2.20%	1316.00	1400.00
	NНз	196	26.69	3.11	11.66%	22.00	30.00
7	CO ₂	196	1369.62	24.14	1.76%	1330.00	1400.00
	NНз	196	27.37	2.60	9.50%	21.00	30.00
8	CO ₂	196	1360.61	27.22	2.00%	1332.00	1400.00
	NНз	196	26.22	3.75	14.30%	20.00	30.00
9	CO ₂	196	1278.49	92.60	7.24%	1185.00	1400.00
	NH3	196	25.68	4.25	16.53%	19.00	30.00

Treatment T1 shows a constant CO2 concentration with a mean of 1400.00 ppm and no variability (SD of 0.00 ppm and CV of 0.00%). This indicates that the treatment applied at the start did not present fluctuations in CO2 emissions. However, the mean CO2 emissions progressively decrease from 1379.55 ppm (T2) to 1278.49 ppm (T9), with increasing variability. The SD varies from 17.24 ppm (T2) to 92.60 ppm (T9), and the CV ranges between 1.25% and 7.24%.

The decrease in CO₂ means across treatments suggests that these interventions are effective in reducing CO₂ concentrations. However, the variability in values, especially in T9 (SD of 92.60 ppm and CV of 7.24%), indicates that this treatment yields results that may be influenced by additional uncontrolled factors or greater heterogeneity in experimental conditions.

Regarding NH₃, the means decrease from 28.45 ppm (T₂) to 25.68 ppm (T₉). The SD ranges between 1.55 ppm (T₂) and 4.25 ppm (T₉), while the CV varies from 5.45% to 16.53%. This reduction suggests an effective role in reducing ammonia concentrations, but variability significantly increases in later treatments, particularly in T₉, which shows the highest CV (16.53%), indicating less consistency in NH₃ emissions under this treatment, potentially linked to fluctuations in experimental conditions or pig responses.

The high variability observed in CO₂ and NH₃ emissions in some treatments, particularly T₉, may be multifactorial, stemming from the interaction between experimental factors (diet variations, physical environment, animal handling), biological factors (changes in intestinal microbiota, individual metabolism), and technical factors (variations in measurement accuracy, sampling frequency, and duration of measurement periods).

Comparison of CO2 and NH3 emission means

The variance analysis yielded a P-value of 0.0001, indicating highly significant differences between treatments in terms of CO₂ emissions. The comparison of mean CO₂ emissions (Figure 1) shows the results for pigs subjected to nine different treatments, which combine various proportions of commercial feed (CF), purple sweet potato flour (PSPF), and *Lactobacillus plantarum* (LP). The bars reflect the mean CO₂ emissions in parts per million (ppm) for each treatment, with letters indicating significant differences between groups.

Treatment T1 (100% CF):

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

T1 presents the highest average CO₂ emission (1410 ppm) and is labeled with the letter "F," indicating it is significantly different and higher compared to other treatments. This suggests that the exclusive use of commercial feed without additions of PSPF or LP leads to the highest CO₂ emissions.

Treatments T2 and T3 (100% CF with addition of 20 mL and 40 mL LP):

The addition of *Lactobacillus plantarum* in T2 and T3 reduces CO2 emissions compared to T1, with averages of 1380 and 1365 ppm, respectively. These treatments are labeled with the letters "E" and "D," indicating that although there is a significant reduction in emissions relative to T1, there is also a significant difference between T2 and T3.

Treatments T4 and T5 (80% CF + 20% PSPF and its combination with LP):

T4, which includes 20% purple sweet potato flour, shows a CO₂ emission mean similar to T₃ (1365 ppm) and is also labeled with the letter "D." The addition of 20 mL of LP to T₄ (forming T₅) results in significantly lower emissions (1325 ppm), labeled with the letter "B." This indicates that the combination of PSPF with a moderate amount of LP has a considerable impact on reducing CO₂ emissions.

Treatments T6, T7, and T8 (80% CF + 20% PSPF with 40 mL LP and variations with 60% CF + 40% PSPF):

T6 (80% CF + 20% PSPF + 40 mL LP) shows a reduction in emissions (1340 ppm) and is labeled "C." This treatment is not significantly different from T7 and T8, which use a higher proportion of PSPF (40%) with varying doses of LP (20 mL and 40 mL). T7 and T8 have similar average emissions (1340 ppm) and share the label "C," suggesting that a higher proportion of PSPF with LP does not offer a significant additional advantage over T6.

Treatment T9 (60% CF + 40% PSPF + 40 mL LP):

This treatment presents the lowest average CO₂ emission (1270 ppm) and is labeled "A," indicating that it is significantly different and lower than all other treatments. The combination of a high proportion of PSPF with the maximum dose of LP is the most effective in reducing CO₂ emissions.

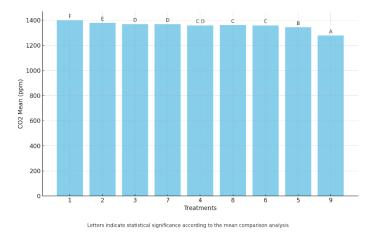


Figure 1. Comparison of mean CO2 emissions across experimental treatments in pigs

In summary, the treatment based solely on commercial feed (T1) results in the highest CO₂ emissions, consistent with trends observed in other studies where conventional diets tend to increase gas production in pigs. Additionally, the inclusion of *Lactobacillus plantarum* reduces CO₂ emissions, especially when combined with purple sweet potato flour. The use of PSPF at both 20% and 40%, combined with LP, shows a clear reduction in emissions

Comparison of ammonia (NH3) emission averages

Figure 2 shows the comparison of ammonia (NH₃) emission averages in pigs subjected to different treatments ($P \le 0.001$), which include combinations of commercial feed (CF), purple sweet potato flour (PSPF), and *Lactobacillus plantarum* (LP). The bars represent the NH₃ emission means in parts per million (ppm) for each treatment, and the letters above the bars indicate significant differences between treatments.

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

Treatment T1 (100% CF):

This treatment presents the highest average NH₃ emission (30 ppm) and is significantly different from the other treatments, as indicated by the letter "D" in the graph. This suggests that the exclusive use of commercial feed (without the addition of PSPF or LP) leads to higher ammonia emissions.

Treatments T2 and T3 (100% CF with 20 mL and 40 mL of LP):

The inclusion of *Lactobacillus plantarum* in the treatments (T2 and T3) significantly reduces NH3 emissions compared to T1, with averages around 28 ppm. Both treatments are labeled with the letter "C," indicating that there is no statistically significant difference between them. This suggests that the addition of LP, regardless of the dose, reduces NH3 emissions.

Treatments T4 and T5 (80% CF + 20% PSPF and its combination with LP):

The inclusion of purple sweet potato flour in the treatment (T4) also shows a reduction in NH3 emissions (28.5 ppm), sharing the letter "C" with T2 and T3, indicating similarity in emission reductions among these treatments.

The addition of 20 mL of LP to the 80% CF + 20% PSPF treatment (T5) results in a greater reduction in NH3 emissions (27 ppm), labeled with the letter "B." This suggests that the combination of PSPF with LP has an additive effect in reducing emissions.

Treatments T6, T7, and T8 (80% CF + 20% PSPF with 40 mL LP and variations with 60% CF + 40% PSPF):

T6 (80% CF + 20% PSPF + 40 mL LP) shows a similar average NH3 emission to T5, and both share the label "A/B." This indicates that there are no significant differences between these treatments, but both are more effective than T1.

Treatments T7 and T8, which use a higher proportion of PSPF (40%) with different levels of LP (20 mL and 40 mL, respectively), show similar average emissions (26 ppm), also labeled "A/B." This reinforces the idea that increasing the proportion of PSPF combined with the addition of LP contributes to the reduction of NH₃.

Treatment T9 (60% CF + 40% PSPF + 40 mL LP):

This treatment shows the lowest NH3 emission (24.5 ppm) and is labeled with the letter "A," indicating that it is significantly different and more effective than most other treatments in reducing NH3 emissions. The combination of a high proportion of PSPF with the maximum dose of LP appears to be the most effective in reducing ammonia emissions.

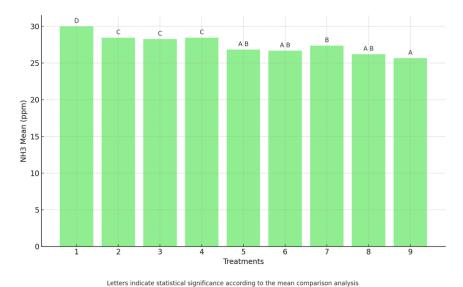


Figure 2. Comparison of NH3 emission means across experimental treatments in pigs

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

The treatment based exclusively on commercial feed (T1) results in the highest NH3 emissions, which is consistent with the idea that the base diet directly influences the amount of ammonia produced.

The inclusion of *Lactobacillus plantarum*, whether at 20 mL or 40 mL, in combination with CF or PSPF results in a significant reduction in NH3 emissions. Additionally, purple sweet potato flour also contributes to the reduction of emissions, especially when used at higher proportions (40% PSPF) and combined with LP.

The T9 treatment, which combines 60% CF with 40% PSPF and 40 mL of LP, is the most effective in reducing NH3 emissions, suggesting a synergistic effect between these components.

This analysis is crucial for understanding how different dietary combinations can influence ammonia emissions, an environmental pollutant, and suggests that the use of ingredients such as purple sweet potato flour and probiotics may be a viable strategy to reduce these emissions in pig production.

Analysis of CO2 and NH3 emissions with and without treatment (Residual Effect)

The descriptive statistics for both phases (with and without treatments) are presented in Table 3. **With treatment**: The CO₂ concentration during the first phase shows a high mean of 1380.23 ppm with a relatively low standard deviation (35.32 ppm), indicating that the values are tightly clustered around the mean. The CV of 2.56% also suggests low relative variability in the measurements, indicating that the treatment effect is consistent in CO₂ production. The NH₃ mean is 27.86 ppm with a standard deviation of 2.67 ppm. The CV of 9.59% indicates higher relative variability compared to CO₂, which may suggest that the treatment affects NH₃ production more diversely. However, the values still fall within a relatively narrow range (20.00 - 30.00 ppm).

Without treatment (residual effect): During the second phase, the CO₂ concentration slightly decreases to a mean of 1330.76 ppm. The standard deviation increases to 54.44 ppm, indicating greater dispersion in the values. The CV of 4.09% suggests higher relative variability compared to the first phase, which may reflect a more variable residual effect of the treatment on CO₂ production. The NH₃ mean during the second phase is slightly lower (27.16 ppm) compared to the first phase. The standard deviation of 3.39 ppm and the CV of 12.47% indicate greater relative variability in NH₃ production during this phase, suggesting that the residual effect of the treatment may be less consistent or influenced by additional factors generating more variability in ammonia emissions.

Table 3. Descriptive Statistics of CO₂ and NH₃ Emissions in Pigs With and Without Treatment of Purple Sweet Potato Flour and *Lactobacillus plantarum*

Group	Variable	n	Mean	SD	CV	Min	Max
With treatment	CO ₂	972	1380.23	35.32	2.56%	1185.00	1400.00
	NH3	972	27.86	2.67	9.59%	20.00	30.00
Without treatment	CO ₂	792	1330.76	54.44	4.09%	1188.00	1400.00
	NH3	792	27.16	3.39	12.47%	19.00	30.00

Group comparison (Phases):

Table 4 presents the results of the Wilcoxon test for independent samples, aiming to compare CO₂ and NH₃ concentrations at two different phases: with treatment application (Group 1) and without treatment application (Group 2). The Wilcoxon test for CO₂ returns a p-value of 0.000001, indicating a highly significant difference between the two phases. This implies that the application of treatments had a considerable influence on CO₂ concentration. For NH₃, the test shows a p-value of 0.0427, indicating a statistically significant difference between the two phases (with and without treatment application) at the 5% significance level. This suggests that the application of treatments had a significant effect on NH₃ concentration.

Table 4. Statistical Results of the Wilcoxon Test for CO₂ and NH₃ Emissions in Pigs With and Without Treatment of Purple Sweet Potato Flour and *Lactobacillus plantarum*

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

Variable	Group 1	Group 2	n (1)	n (2)	Mean (1)	Mean (2)	SD (1)	SD (2)	w	p (2 tails)
CO ₂	1	2	972	792	1380.23	1330.76	35.32	54.44	44996250	0.000001
NH3	1	2	972	792	27.86	27.16	2.67	3.39	67864400	0.0427

Group 1: With treatment; Group 2: Without treatment.

The Wilcoxon test for CO₂ returns a p-value of 0.000001, indicating a highly significant difference between the two groups (phases). This implies that the application of treatments had a considerable influence on CO₂ concentration. For NH₃, the test shows a p-value of 0.0427, indicating a statistically significant difference between the two groups (with and without treatment application) at the 5% significance level. This suggests that the application of treatments had a significant effect on NH₃ concentration.

The results of the Wilcoxon test suggest that both CO₂ and NH₃ concentrations were significantly affected by the application of treatments. The statistical significance found in both variables reinforces the importance of the applied treatments in modifying the concentrations of these gases.

DISCUSSION

Global impact of emissions in pig production

Pig production is responsible for approximately 668 million tons of CO₂-eq annually, representing 9% of emissions from the livestock sector (Gerber et al., 2013). In addition to carbon dioxide (CO₂), other gases such as ammonia (NH₃), methane (CH₄), volatile organic compounds (VOCs), and hydrogen sulfide (H₂S) contribute to environmental problems such as eutrophication, reduced air quality, and climate change (Dalby et al., 2020). In response to this challenge, the FAO (2023) highlights the need for strategies to reduce greenhouse gas emissions, particularly considering the projected increase in demand for animal products by 2050.

Gas emissions and factors influencing them

CO₂ emissions in pigs are influenced by physiological stage, body weight, and food intake. In the case of CH₄, its production is primarily related to fermentation in the hindgut, underscoring the importance of diet in generating these gases. CO₂, CH₄, and N₂O emissions represent 81%, 17%, and 2% of total emissions in pig farms, respectively (Philippe & Nicks, 2015).

Effect of a purple sweet potato and probiotic diet on emissions

The results of this study show that the inclusion of purple sweet potato (*Ipomoea batatas*) and *Lactobacillus plantarum* in pig diets significantly reduces CO₂ and NH₃ emissions. This aligns with previous research highlighting the role of probiotics and phytochemicals in modulating the gut microbiota (Biswas et al., 2024; Vadopalas et al., 2020). This reduction suggests that the polyphenols in purple sweet potato play a key role in improving the conditions for fiber fermentation, thereby reducing the production of harmful gases (Kilua et al., 2019).

Benefits of polyphenols and anthocyanins from purple sweet potato

The anthocyanins in purple sweet potato have demonstrated prebiotic properties, promoting the growth of beneficial bacteria such as *Bifidobacterium* and *Lactobacillus* while inhibiting pathogens like *Staphylococcus aureus* and *Salmonella typhimurium* (Lippolis et al., 2023; Sun et al., 2018; Zhang et al., 2016). These prebiotic effects are crucial in reducing gas emissions by promoting a healthy gut microbiota (Peng et al., 2020).

Reduction of ammonia through dietary fiber and probiotics

The inclusion of dietary fiber in pig diets has a significant impact on manure composition and NH3 and H2S emissions. Studies have shown a 30% reduction in NH3 emissions and a 17% reduction in H2S emissions with high-fiber diets (Trabue et al., 2022). Additionally, better alignment of dietary intake with the physiological needs of pigs also reduces NH3 emissions (Philippe et al., 2011).

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

Probiotics, when combined with prebiotics, can improve digestibility and reduce harmful gas emissions (Chu et al., 2011). In pigs fed diets fermented with *Lactobacillus*, there has been demonstrated improvement in growth performance and reduced fecal and urinary nitrogen excretion (Liu et al., 2024; Lan et al., 2017).

Synergy between polyphenols and probiotics: implications for environmental sustainability

The results of this study confirm that the combination of polyphenols in purple sweet potato and *Lactobacillus plantarum* produces a synergistic effect that significantly contributes to reducing CO₂ and NH₃ emissions in pig production. This interaction between phytochemicals and probiotics favorably modulates gut microbiota, reducing the production of pollutant gases, which is consistent with previous studies (Zhang et al., 2019). These findings have important implications for developing sustainable feeding strategies that promote the use of natural additives, which not only improve animal health but also mitigate environmental impact. Further evaluation of this synergy in various production contexts is recommended to maximize its effect on the sustainability of animal production.

CONCLUSIONS

Based on the results obtained, this study provides evidence of the positive influence of feed additives, specifically polyphenols and probiotics, in pig diets, highlighting the use of purple sweet potato (*Ipomoea batatas L.*) and *Lactobacillus plantarum*. These additives contribute to the reduction of CO₂ and NH₃ emissions, suggesting their potential to mitigate the environmental impact associated with pig production.

It is essential to conduct further evaluations that consider the effects of this combination of additives on productive parameters and animal health. A deeper understanding of the interaction between animal health status and feed additives could facilitate the development of sustainable strategies that promote the reduction of harmful gases for the environment.

REFERENCES

- [1] Bindelle, J., Buldgen, A., Delacollette, M., Wavreille, J., Agneessens, R., Destain, J. P., & Leterme, P. (2009). Influence of source and concentrations of dietary fiber on in vivo nitrogen excretion pathways in pigs as reflected by in vitro fermentation and nitrogen incorporation by fecal bacteria. *Journal of animal science*, 87(2), 583-593. https://doi.org/10.2527/jas.2007-0717
- [2] Chu, G. M., Lee, S. J., Jeong, H., & Lee, S. S. (2011). Efficacy of probiotics from anaerobic microflora with prebiotics on growth performance and noxious gas emission in growing pigs. *Animal Science Journal = Nihon Chikusan Gakkaiho*, 82(2), 282-290. https://doi.org/10.1111/j.1740-0929.2010.00828.x
- [3] Dalby, F. R., Svane, S., Sigurdarson, J. J., Sørensen, M. K., Hansen, M. J., Karring, H., & Feilberg, A. (2020). Synergistic tannic acid-fluoride inhibition of ammonia emissions and simultaneous reduction of methane and odor emissions from livestock waste. *Environmental Science & Technology*, 54(12), 7639-7650. https://doi.org/10.1021/acs.est.0c01231
- [4] Gasaly, N., & Gotteland, M. (2022). Interference of dietary polyphenols with potentially toxic amino acid metabolites derived from the colonic microbiota. *Amino Acids*, 311-324. https://doi.org/10.1007/s00726-021-03034-3
- [5] Gerber, P. J., Steinfeld, H., Henderson, B., Mottet, A., Opio, C., Dijkman, J., Falcucci, A., & Tempio, G. (2013). Enfrentando el cambio climático a través de la ganadería. https://openknowledge.fao.org/server/api/core/bitstreams/57306995-b497-442a-a127-81a890612f34/content/i3437s.htm
- [6] Guo, F., & Shahidi, F. (2023). Can anthocyanins replace antibiotics in food and animal feed? A review. *Trends in Food Science & Technology*. https://doi.org/10.1016/j.tifs.2023.104219
- [7] Hossain, M. M., Cho, S. B., & Kim, I. H. (2024). Strategies for reducing noxious gas emissions in pig production: A comprehensive review on the role of feed additives. *Journal of Animal Science and Technology*, 66(2), 237. https://doi.org/10.5187/jast.2024.e15
- [8] Hurtado, E. A., Farfan, C. L. F., Cheme, K. L. M., & Marcillo, G. A. C. (2024). African Journal of Biological Sciences. Afr.J.Bio.Sc. 6(8). https://www.afjbs.com/uploads/paper/f561e95bae5cce9e859861ae7531a82a.pdf

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

- [9] Kilua, A., Nomata, R., Nagata, R., Fukuma, N., Shimada, K., Han, K. H., & Fukushima, M. (2019). Purple sweet potato polyphenols differentially influence the microbial composition depending on the fermentability of dietary fiber in a mixed culture of swine fecal bacteria. *Nutrients*, 11(7), 1495. https://doi.org/10.3390/nu11071495
- [10] Lan, R., Tran, H., & Kim, I. (2017). Effects of probiotic supplementation in different nutrient density diets on growth performance, nutrient digestibility, blood profiles, fecal microflora, and noxious gas emission in weaning pig. *Journal of the Science of Food and Agriculture*, *97*(4), 1335-1341. https://doi.org/10.1002/jsfa.7871
- [11] Lippolis, T., Cofano, M., Caponio, G. R., De Nunzio, V., & Notarnicola, M. (2023). Bioaccessibility and bioavailability of diet polyphenols and their modulation of gut microbiota. *International Journal of Molecular Sciences*, 24(4), 3813. https://doi.org/10.3390/ijms24043813
- [12] Liu, H., Wang, S., Chen, M., Ji, H., & Zhang, D. (2024). Effects of Lactobacillus-fermented low-protein diets on the growth performance, nitrogen excretion, fecal microbiota, and metabolomic profiles of finishing pigs. *Scientific Reports*, 14(1), 8612. https://doi.org/10.1038/s41598-024-58832-y
- [13] Peng, M., Tabashsum, Z., Anderson, M., Truong, A., Houser, A. K., Padilla, J., Akmel, A., Bhatti, J., Rahaman, S. O., & Biswas, D. (2020). Effectiveness of probiotics, prebiotics, and prebiotic-like components in common functional foods. *Comprehensive Reviews in Food Science and Food Safety*, 19, 1908–1933. https://doi.org/10.1111/1541-4337.12565
- [14] Philippe, F. X., & Nicks, B. (2015). Review on greenhouse gas emissions from pig houses: Production of carbon dioxide, methane and nitrous oxide by animals and manure. *Agriculture, Ecosystems & Environment, 199*, 10-25. https://doi.org/10.1016/j.agee.2014.08.015
- [15] Philippe, F. X., Cabaraux, J. F., & Nicks, B. (2011). Ammonia emissions from pig houses: Influencing factors and mitigation techniques. *Agriculture, Ecosystems & Environment,* 141(3-4), 245-260. https://doi.org/10.1016/j.agee.2011.03.012
- [16] Rowland, I., Gibson, G., Heinken, A., Scott, K., Swann, J., Thiele, I., & Tuohy, K. (2018). Gut microbiota functions: Metabolism of nutrients and other food components. *European Journal of Nutrition*, *57*, 1-24. https://doi.org/10.1007/s00394-017-1445-8
- [17] Serra, V., Salvatori, G., & Pastorelli, G. (2021). Dietary polyphenol supplementation in food producing animals: Effects on the quality of derived products. *Animals*, 11(2), 401. https://doi.org/10.3390/ani11020401
- [18] Smith, P., Martino, D., & Cai, Z. (2008). Greenhouse gas mitigation in agriculture. *Philosophical Transactions of the Royal Society B: Biological Sciences*, *363*, 789-813.
- [19] Sun, H., Zhang, P., Zhu, Y., Lou, Q., & He, S. (2018). Antioxidant and prebiotic activity of five peonidin-based anthocyanins extracted from purple sweet potato (*Ipomoea batatas* (L.) Lam.). *Scientific Reports*, 8(1), 5018. https://doi.org/10.1038/s41598-018-23397-0
- [20] Trabue, S. L., Kerr, B. J., Scoggin, K. D., Andersen, D. S., & van Weelden, M. (2022). Swine diets: Impact of carbohydrate sources on manure characteristics and gas emissions. *Science of The Total Environment*, 825, 153911. https://doi.org/10.1016/j.scitotenv.2022.153911
- [21] Tufarelli, V., Crovace, A. M., Rossi, G., & Laudadio, V. (2017). Effect of a dietary probiotic blend on performance, blood characteristics, meat quality and faecal microbial shedding in growing-finishing pigs. *South African Journal of Animal Science*, 47, 875-882. https://hdl.handle.net/10520/EJC-af36a9892
- [22] Vadopalas, L., Ruzauskas, M., Lele, V., Starkute, V., Zavistanaviciute, P., Zokaityte, E., Zokaityte, E., Bartkevics, V., Badaras, S., Klupsaite, D., Mozuriene, E., Dauksiene, A., Sidlauskiene, S., Gruzauskas, R., & Bartkiene, E. (2020). Pigs' feed fermentation model with antimicrobial lactic acid bacteria strains combination by changing extruded soya to biomodified local feed stock. *Animals*, 10(5), 783. https://doi.org/10.3390/ani10050783
- [23] Vidal, A. R., Zaucedo-Zuñiga, A. L., & de Lorena Ramos-García, M. (2018). Propiedades nutrimentales del camote (*Ipomoea batatas* L.) y sus beneficios en la salud humana. *Revista Iberoamericana de Tecnología Postcosecha*, 19(2). https://www.redalvc.org/journal/813/81357541001/81357541001.pdf
- [24] Vorobel, M., Klym, O., & Kaplinskyi, V. (2024). The effectiveness of the influence on the emission of harmful gases from pig manure during storage in lagoons. *Animal Science Journal. Serie D. Ciencia Animal*, 67(1). https://www.animalsciencejournal.usamv.ro/pdf/2024/issue 1/Art44.pdf
- [25] Zhang, X., Yang, Y., Wu, Z., & Weng, P. (2016). The modulatory effect of anthocyanins from purple sweet potato on human intestinal microbiota in vitro. *Journal of agricultural and food chemistry*, 64(12), 2582-2590. https://doi.org/10.1021/acs.jafc.6b00586

2025, 10(44s) e-ISSN: 2468-4376

https://www.jisem-journal.com/

Research Article

[26] Zhang, Z., Zhang, R., Xiao, H., Bhattacharya, K., Bitounis, D., Demokritou, P., & McClements, D. J. (2019). Development of a standardized food model for studying the impact of food matrix effects on the gastrointestinal fate and toxicity of ingested nanomaterials. *NanoImpact*, 13, 13-25. https://doi.org/10.1016/j.impact.2018.11.002